

## PlexTaq® 5x qPCR Multiplex Master Mix

#4000, Manual Version 5, January 25, 2021

Store at -20°C

### Contents

PlexTaq® 5x qPCR Multiplex Master Mix contains all components necessary for rapid, sensitive and reproducible quantification of DNA and cDNA. An engineered DNA polymerase and an optimized buffer including ultrapure dNTPs are key components of the ready to use mix. A hot-start formulation of the included DNA polymerase prevents false amplification during the reaction set-up.

### Description

PlexTaq® 5x qPCR Multiplex Master Mix is a ready to use reaction mix. It contains all components necessary for a successful and reliable probe based qPCR in all standard real-time PCR cyclers. Only primers, template and a fluorescence-based hydrolysis probe need to be added.

This mix provides robust PCR performance for a wide range of qPCR applications. The buffer is optimized to function with a wide range of templates including human-, mammal-, and plant-derived samples. The PlexTaq® 5x qPCR Multiplex Master Mix ensures reproducible results, significantly reduces set-up times and the risk of pipetting mistakes.

### Recommendations for PCR/ Reaction Setup

#### PCR Mix

Component	Volume	Final concentration
PlexTaq® 5x qPCR Multiplex Master Mix	5 µl	1x
Primer forward (10 µM)*	0.5 µl	0.2 µM (0.05-1 µM)
Primer reverse (10 µM)*	0.5 µl	0.2 µM (0.05-1 µM)
Probe	x µl	0.2 µM (0.05-0.3 µM)
Template	x µl	<300 ng** DNA
Nuclease-free water		up to 25 µl total volume

\* Primers should ideally have a GC content of 40-60% typically. For optimal results we recommend amplicon lengths in the range of 60 to 300 bp.

\*\*Suggested template concentration should be about 1 ng - 300 ng (genomic DNA) or 1 ng - 1 pg (plasmid/viral DNA).

#### Typical 2-step PCR protocol

Initial denaturation	95°C	2 min	
Denaturation	95°C	15 sec**	
Annealing/Extension*	60°C	60 sec**	25-40 cycles

\* Typically, the annealing temperature is about 3-5°C below the calculated melting temperature of the primers used.

\*\* Suggested cycling times depend strongly on the cycler, template and amplicon length. For some probe systems a separate annealing and extension steps may be necessary.

Please notice that an initial denaturation of more than 2 min is not needed. Nevertheless, an extended initial denaturation of up to 15 min has no negative effect on the PCR performance.

PCR protocol times and temperatures may vary depending on the used cycler, the nature of template and the amplicon length.

### Cycler and Probe compatibility

This product is compatible for the use with any probe system and qPCR cycler not requiring a passive reference dye.

### Quality Control Assays

PlexTaq® 5x qPCR Multiplex Master Mix is tested in standard qPCR. The product demonstrates linearity of amplification over a specified serial dilution of human genomic DNA. The activity of DNA polymerase is monitored and adjusted to a specific DNA polymerase activity using an artificial DNA template and a DNA primer. Enzyme concentration is determined by protein-specific staining. Please inquire more information at info@mypols.de for the lot-specific concentration. No contamination has been detected in standard test reactions.

### Material Safety Data (MSDS)

According to OSHA 29CFR1910.1200, Australia [NOHSC:1005, 1008 (1999)] and the EU Directives 67/548/EC, 1999/45/EC and 1272/2008 (CLP Regulation) any products which do not contain more than 1% of a component classified as dangerous or hazardous nor more than 0.1% of a component classified as carcinogenic, do not require a MSDS. However, we recommend the use of gloves, lab coats and eye protection when working with these or any other chemical reagents. myPOLS Biotec takes no liability for damage resulting from handling or contact with this product. This product is not hazardous, not toxic, not IATA-restricted. Product is not from human, animal or plant origin. The source of the product is recombinant protein expression in *E. coli*. The product is for research use only and may be used for *in-vitro* experiments only.

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### Important notes

- Thaw and keep all reagents on ice.
- A precipitate might form after storage at -20°C. Vortex the mix until the precipitate is gone prior to use.
- Always include a control without template.
- Primers should ideally have a GC content of 40-60% typically. For optimal results we recommend amplicon lengths in the range of 60 to 300 bp.
- Minimize the number of freeze-thaw cycles by storing the product in aliquots. For a day-to-day use, we recommend keeping an aliquot at 4°C.
- We recommend the use of disposable gloves, DNase and RNase free filter tips and plastics.

### Troubleshooting

*How can I optimize the PCR conditions and prevent false amplification?*

- The annealing/extension temperature can usually be optimized. Try a temperature gradient and determine the best temperature, which results in a high amplification signal.
- Shorten the extension and annealing time - too long and too many cycles may lead to over-amplification and side-products.

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